



# Flow Cytometry

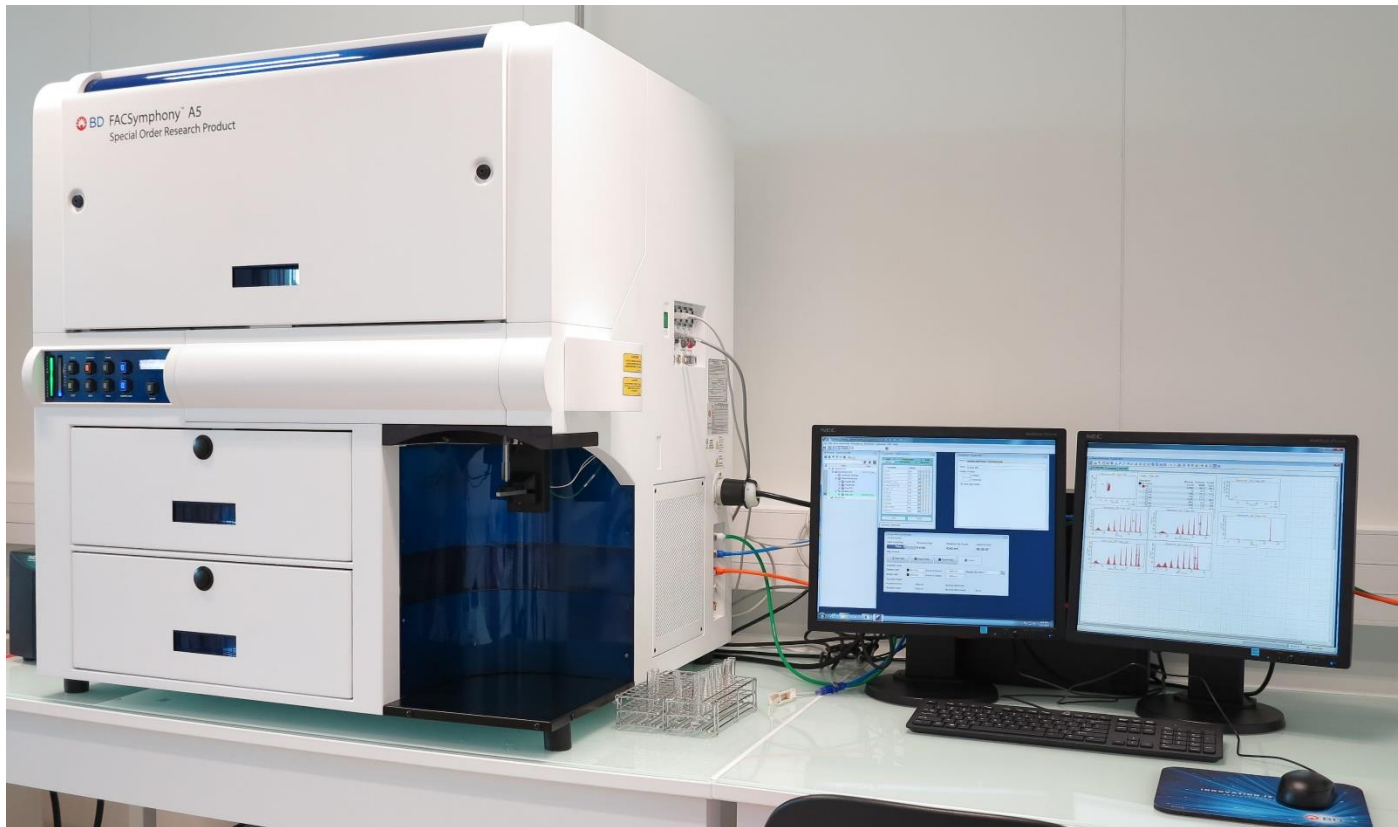
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Medical Biotechnology

Tehran University of Medical Sciences

# “Flow cytometry”: What stands for?

- Measurement (metry) of cells (cyto) in a fluid (flow)



# Flow cytometry applications

- Cell size
- Cell shape
- DNA, RNA and protein content
- Internal and external receptors
- Cell membrane structure
- Apoptosis
- Calcium flux

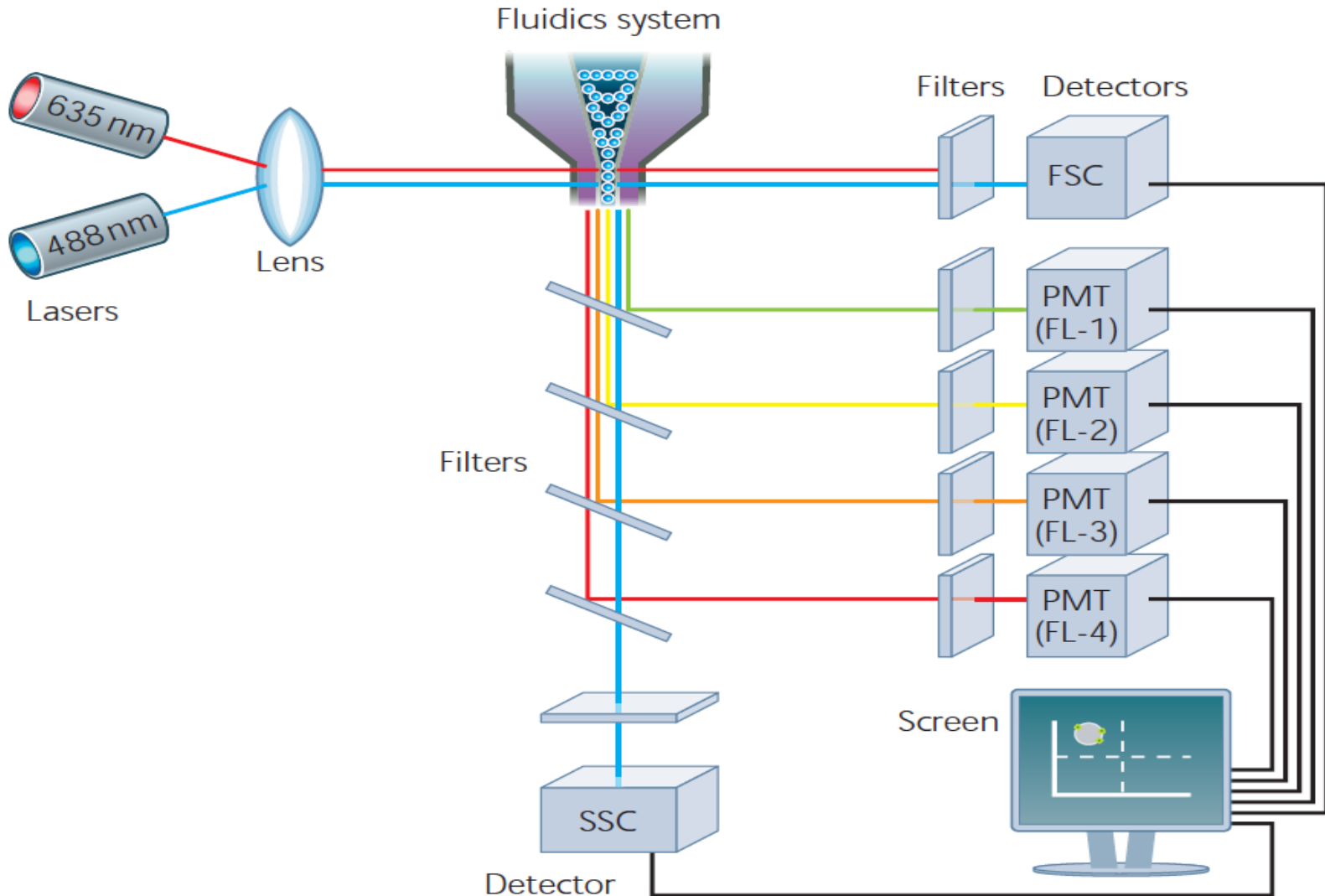
# Advantages

- It is very **FAST**: you can measure tens of thousands of particles per second.
- There is a **LOT of information** to be gained. Many physical and fluorescent parameters are measured simultaneously allowing for the resolution of distinct populations and cellular functions in complex and heterogeneous mixes.
- Data is **qualitative, quantitative** and unbiased.
- **Pure, sterile, live cells** of interest can be retrieved for downstream applications such as culture, functional assays, transplant, imaging and “omics” including DNA and RNA sequencing, proteomics, metabolomics, etc...

# Main parts

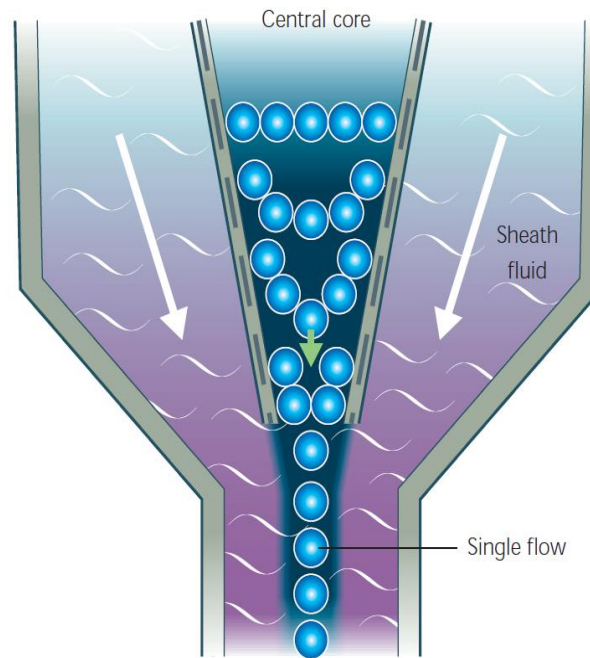
- Fluidic system
- Optical system
- Electronic parts

# Typical Flow cytometer set up



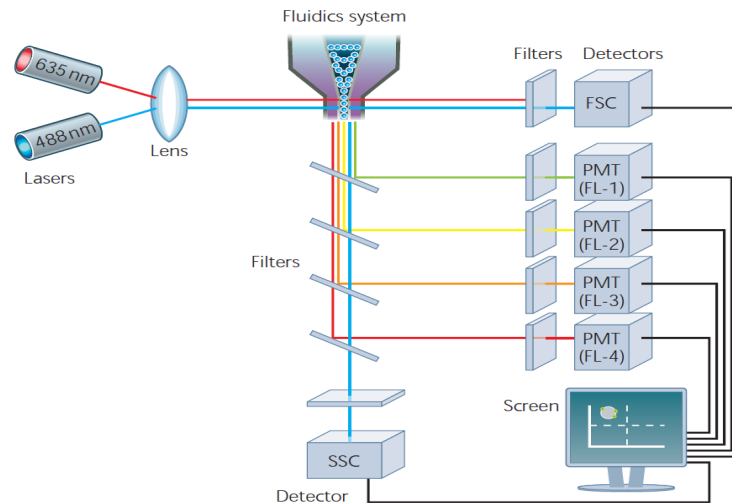
# Fluidic system

- What function for “hydrodynamic focusing”?



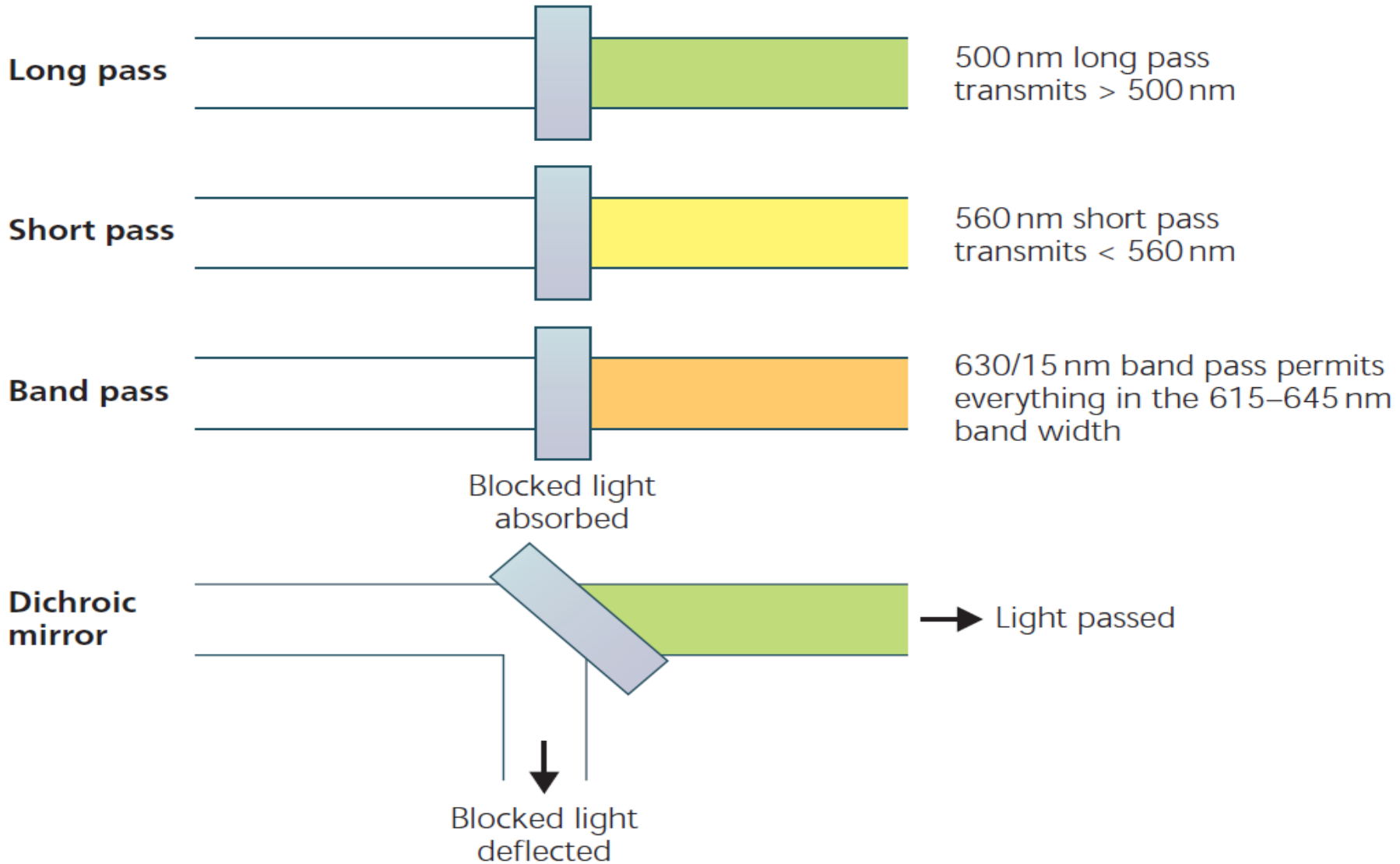
# Optics and detection system

- **Light source:** Laser or arc lamp
- **Lens and filters:**
- **Detectors:** either silicon photodiodes or photomultiplier tubes (PMTs)





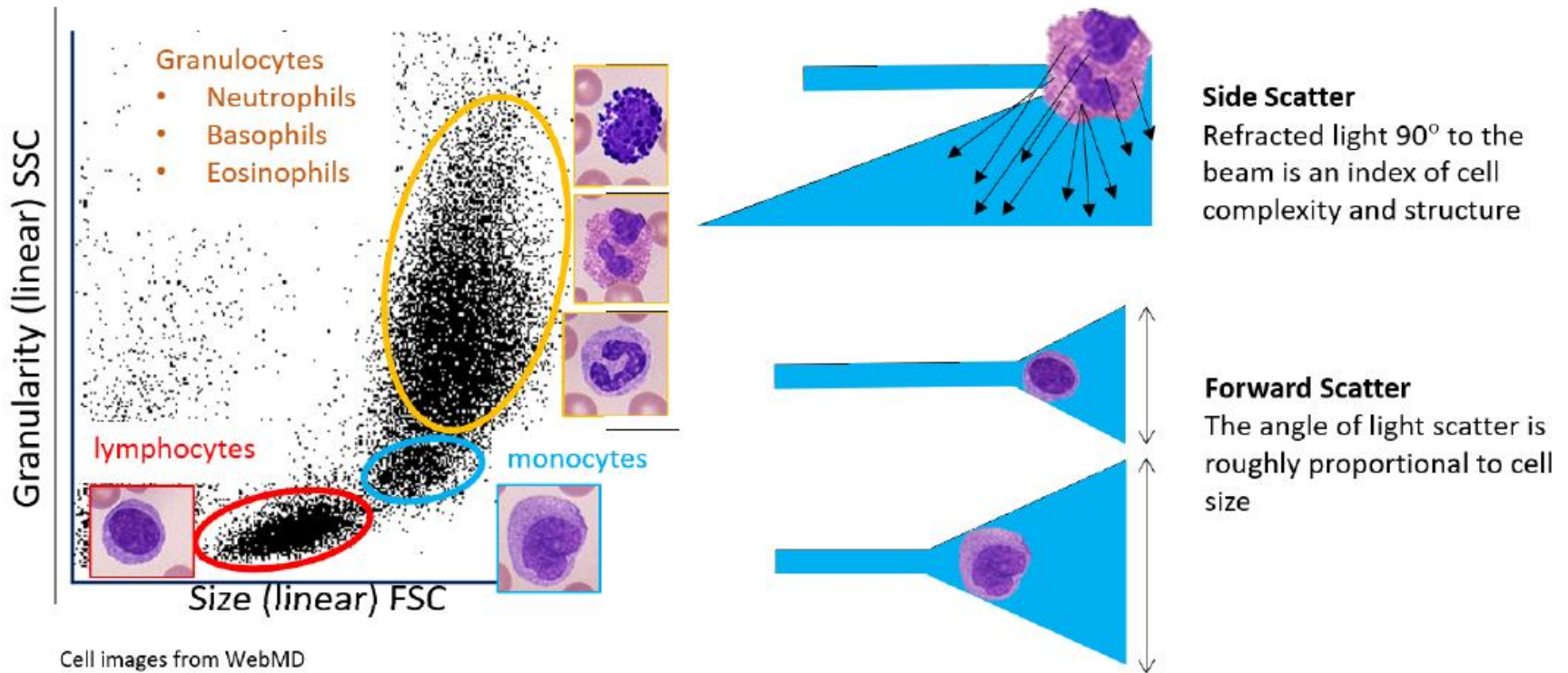
# Types of filters



# Signal Processing

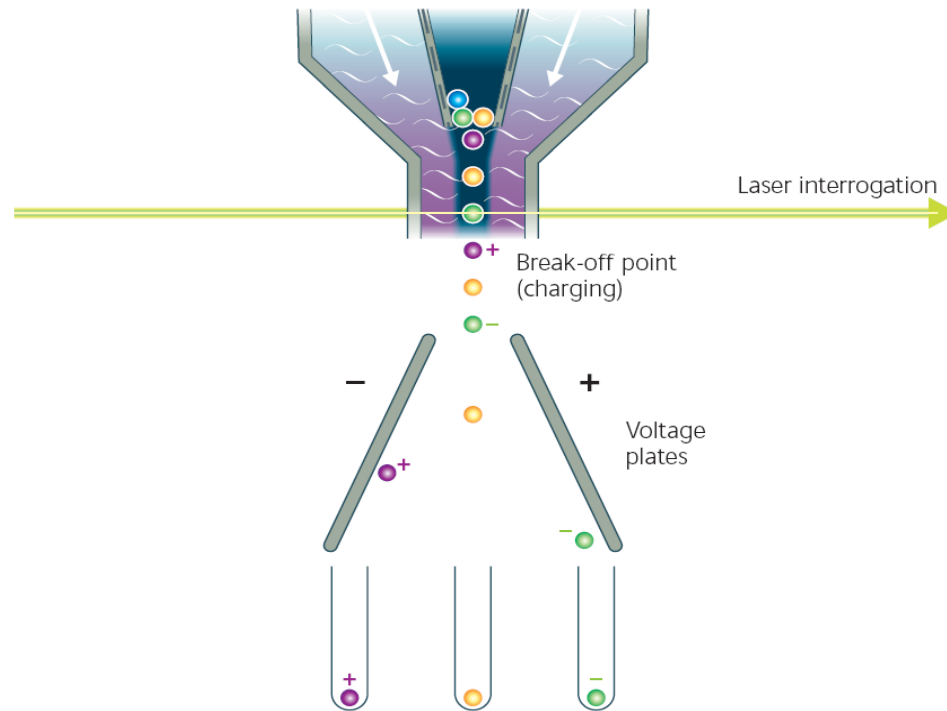
- Log and linear scaling
- **Parameter:** The measurement from each detector, forward scatter (FSC), side scatter (SSC) or fluorescence (FL).
- **Event:** The data acquired in each parameter, refer to the number of cells displaying the physical feature or marker of interest

# Forward Scatter Channel(FSC) and Side Scattering Channel (SSC)



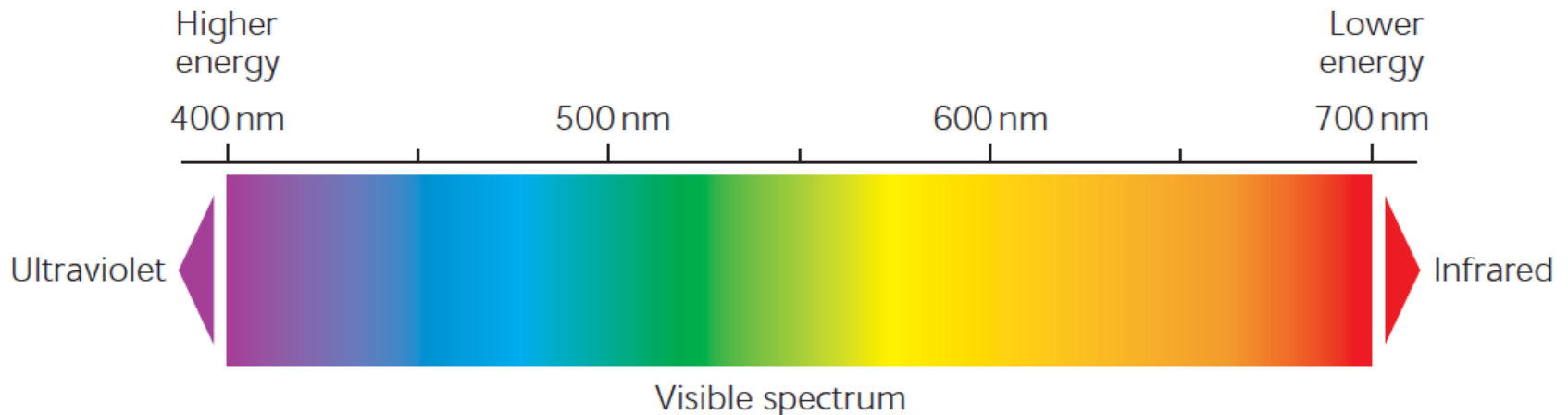
# Cell sorting

- FACS™
- Break-off point

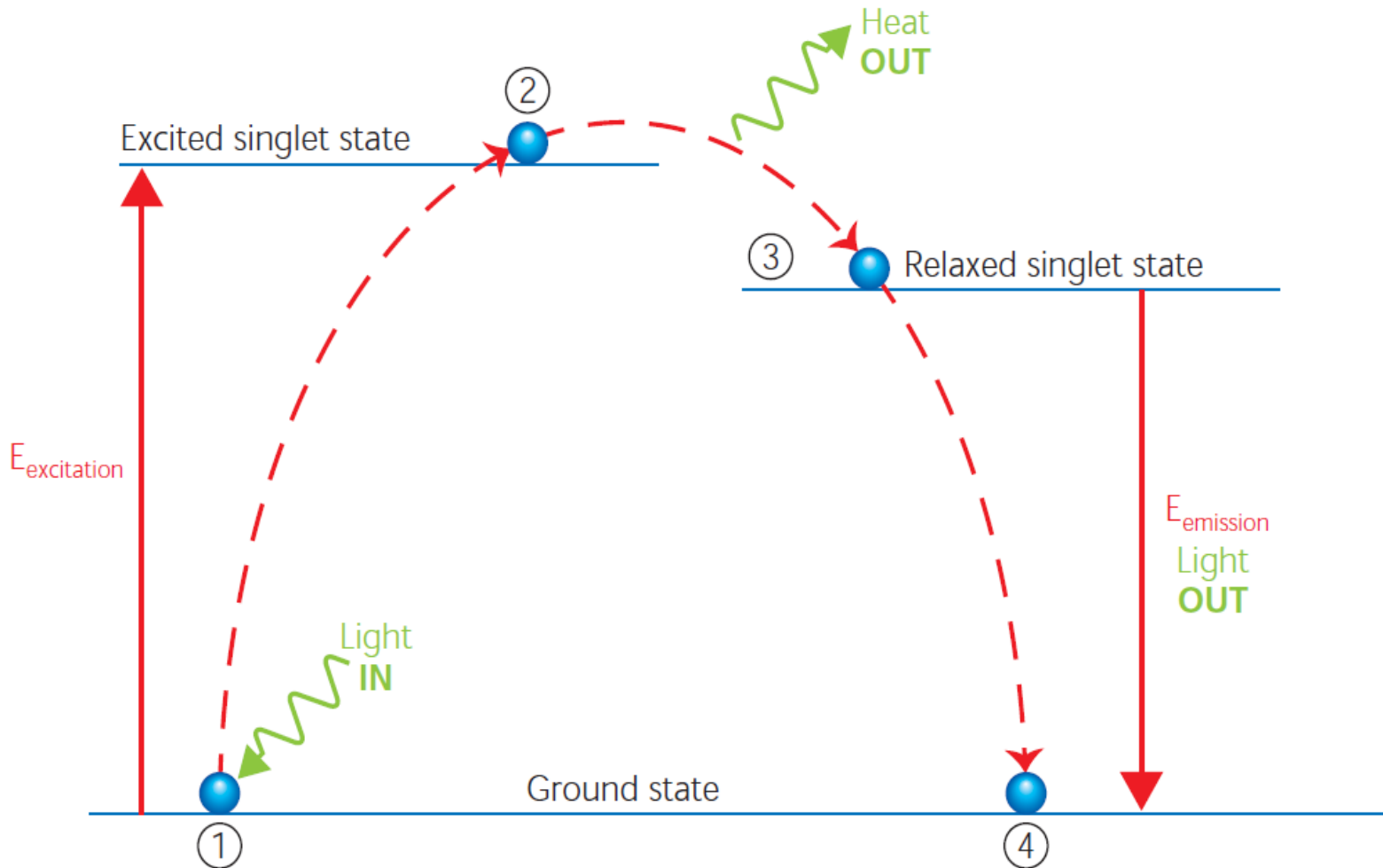


# Principles of fluorescence

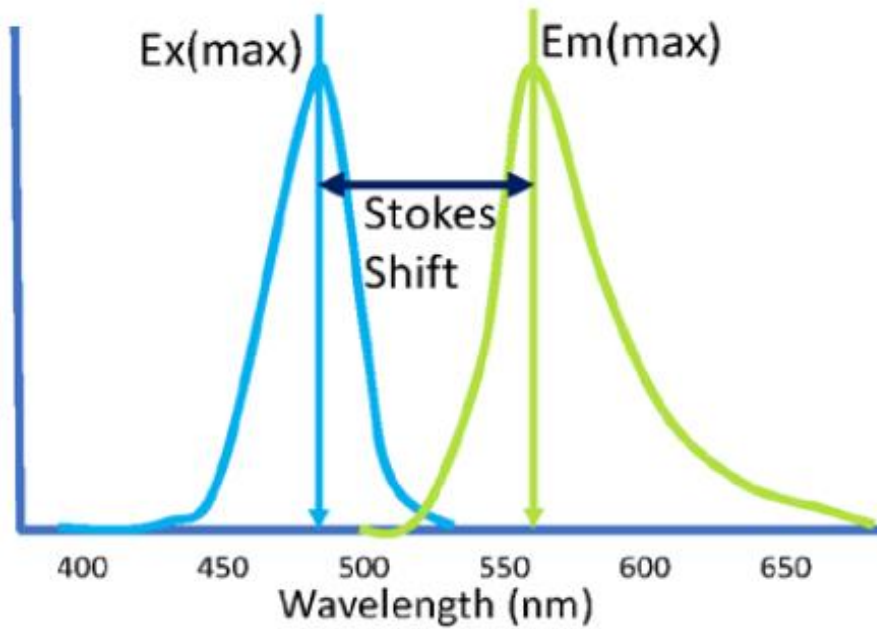
- Fluorochromes: dyes, which accept light energy at a given wavelength and re-emit it at a longer wavelength.
- Excitation and emission
- Light: frequency and wavelength



# Stokes shift

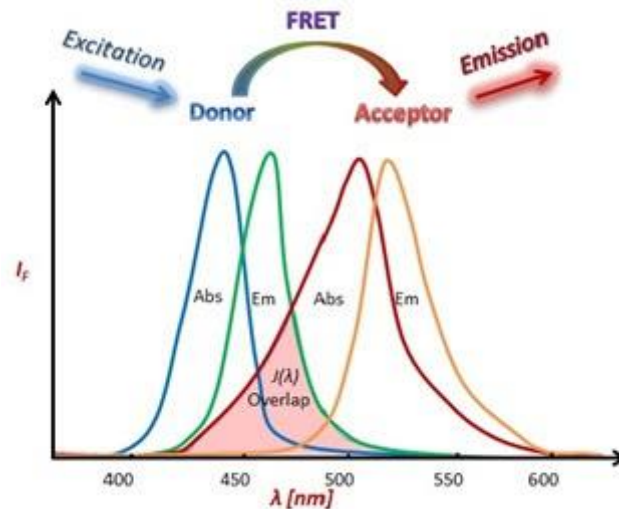


# Stokes shift



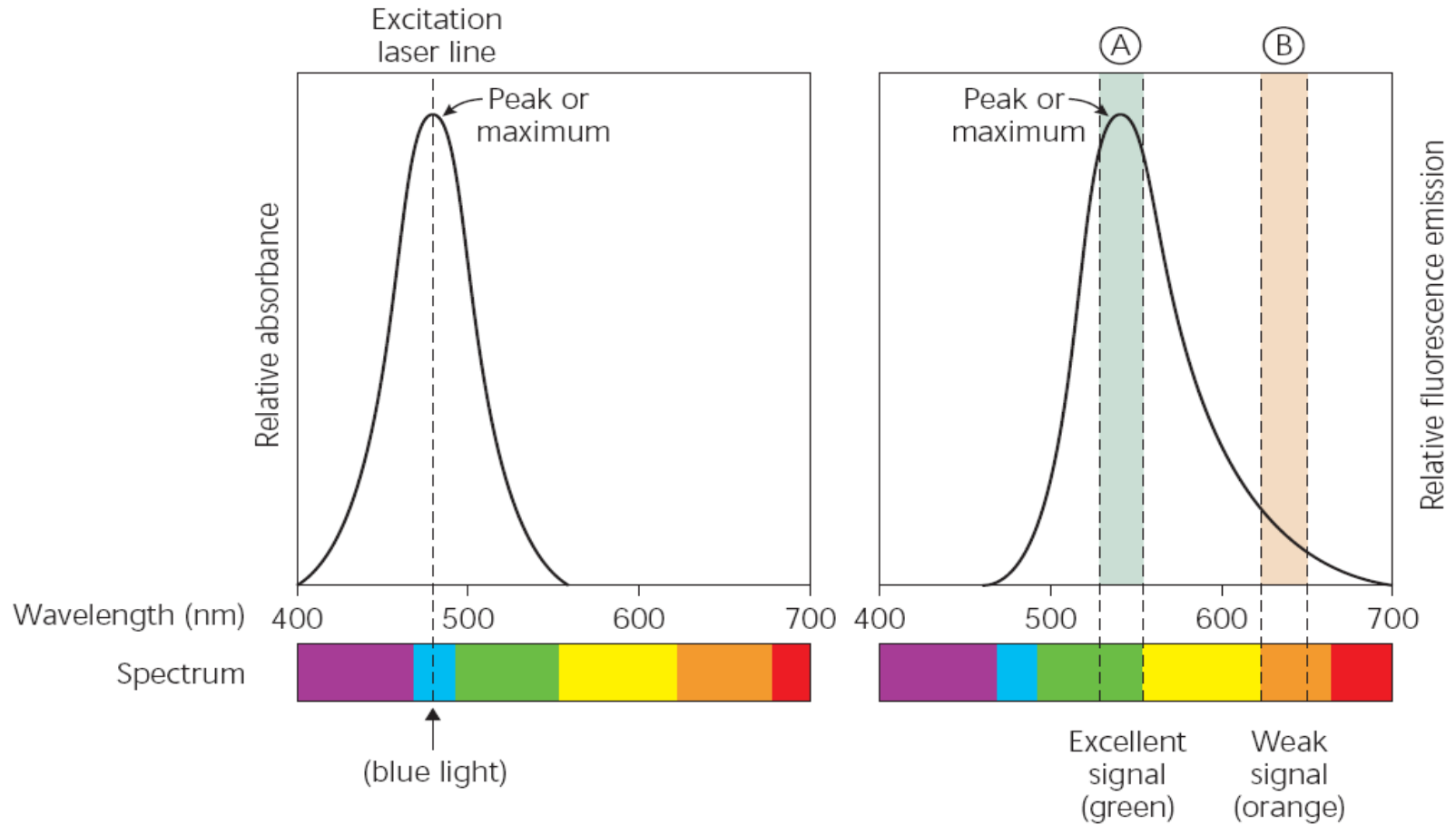
# Importance of Stokes shift

- Single dye vs Tandem dye: FRET (fluorescence resonance energy transfer): a clever way to achieve higher Stokes Shifts and, therefore, increase the number of colors that can be analyzed from a single laser wavelength.

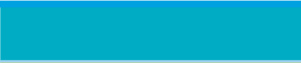






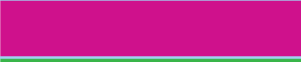








# Maximal absorbance, maximal emission


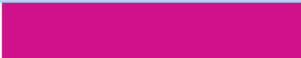



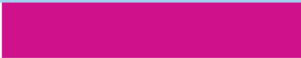




## Single dyes

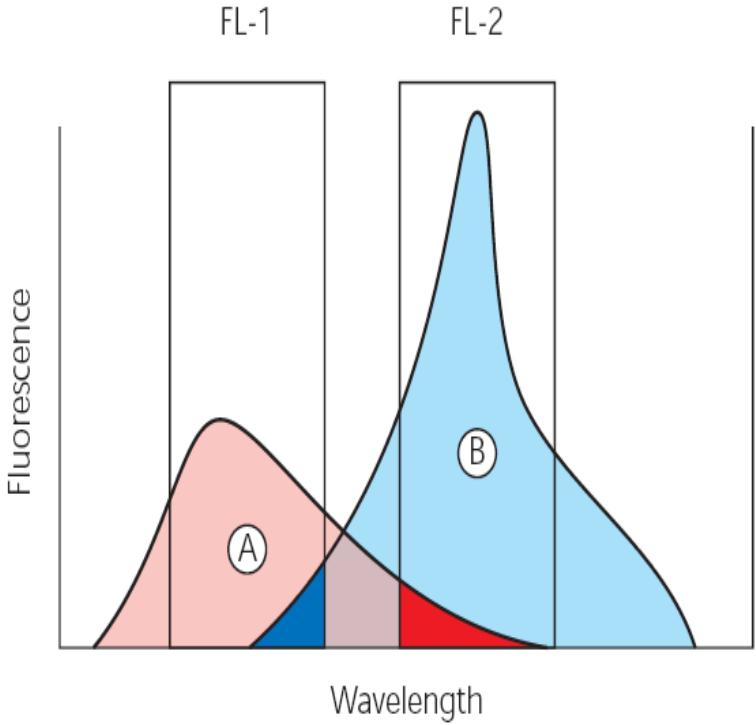
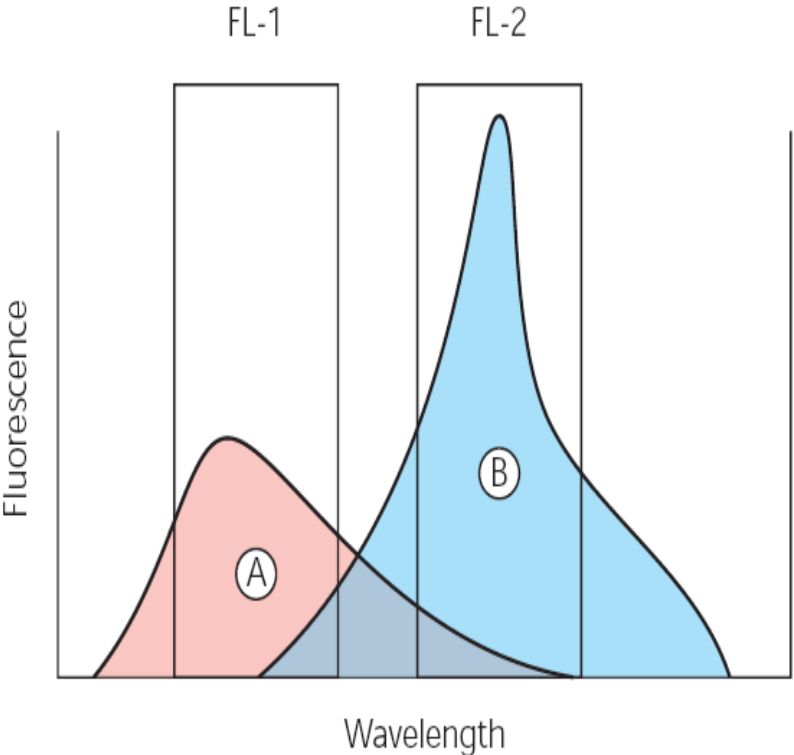
Dye	Laser excitation line (nm)	Maximal absorbance (nm)	Maximal emission (nm)	Fluorescence color
<b>Alexa Fluor<sup>®</sup> 405</b>	405, 407	401	421	
<b>Alexa Fluor<sup>®</sup> 430</b>	405, 407	433	541	
<b>Alexa Fluor<sup>®</sup> 488</b>	488	495	519	
<b>Alexa Fluor<sup>®</sup> 633</b>	633, 635, 647	632	647	
<b>Alexa Fluor<sup>®</sup> 647</b>	633, 635, 647	650	665	
<b>Alexa Fluor<sup>®</sup> 660</b>	633, 635, 647	663	690	
<b>Alexa Fluor<sup>®</sup> 680</b>	633, 635, 647	679	702	
<b>Alexa Fluor<sup>®</sup> 700</b>	633, 635, 647	702	723	Infrared
<b>APC</b>	633, 635, 647	650	661	
<b>FITC</b>	488	490	525	
<b>Pacific Blue<sup>™</sup></b>	405, 407	410	455	
<b>PerCP</b>	488	490	675	
<b>Phycoerythrin</b>	488	490, 565	578	

# FRET (fluorescence resonance energy transfer)

## Tandem dyes

Dye	Laser excitation line (nm)	Maximal absorbance (nm)	Maximal emission (nm)	Fluorescence color
APC-Alexa Fluor® 750	633, 635, 647	650	779	Infrared
APC-Cy5.5	633, 635, 647	650	695	
APC-Cy7	633, 635, 647	650	785	Infrared
PerCP-Cy5.5	488	496, 546	695	
PE-Alexa Fluor® 610	488	496, 546	627	
PE-Alexa Fluor® 647	488	496, 546	667	
PE-Alexa Fluor® 680	488	496, 546	702	
PE-Alexa Fluor® 700	488	496, 546	723	Infrared
PE-Alexa Fluor® 750	488	496, 546	779	Infrared
PE-Cy5.5	488	496, 546	695	
PE-Cy5	488	496, 546	667	
PE-Cy7	488	496, 546	785	Infrared
PE-Texas Red®	488	496, 546	615	

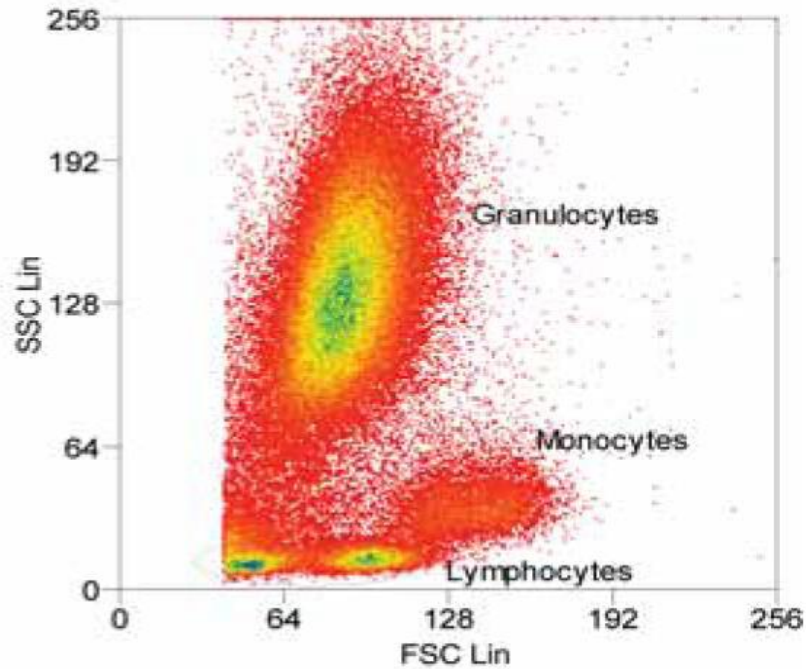
# Fluorescence compensation



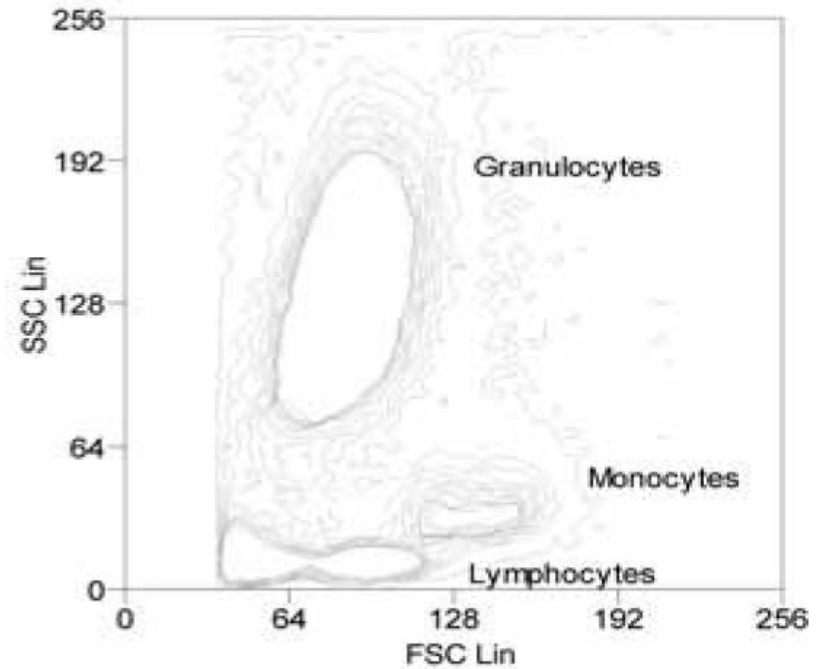
# Data analysis

- Gating: e.g. according to physical characteristics

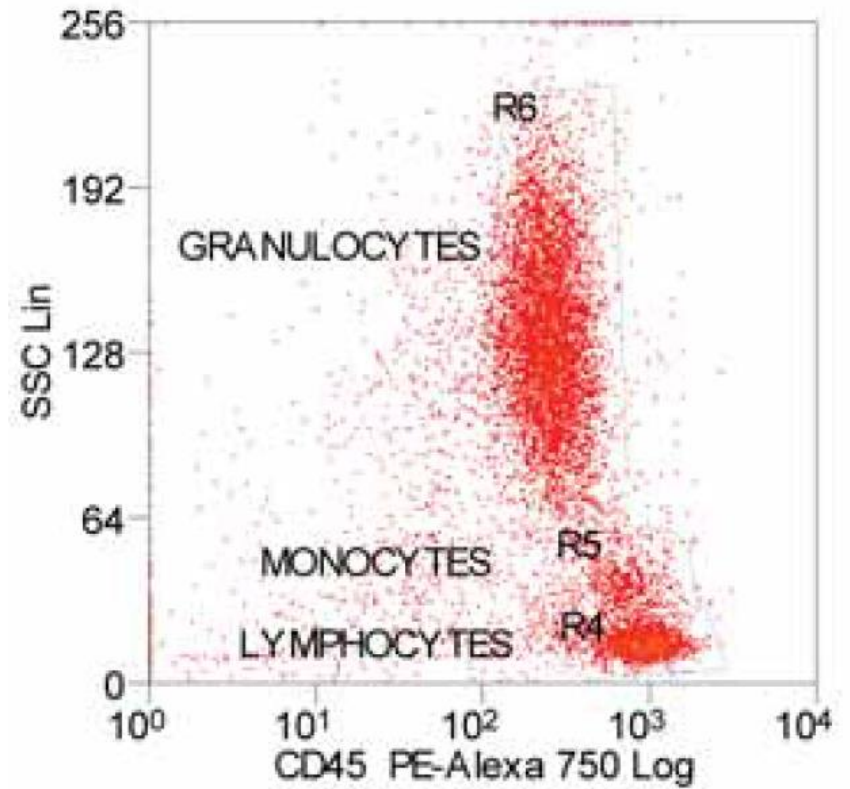
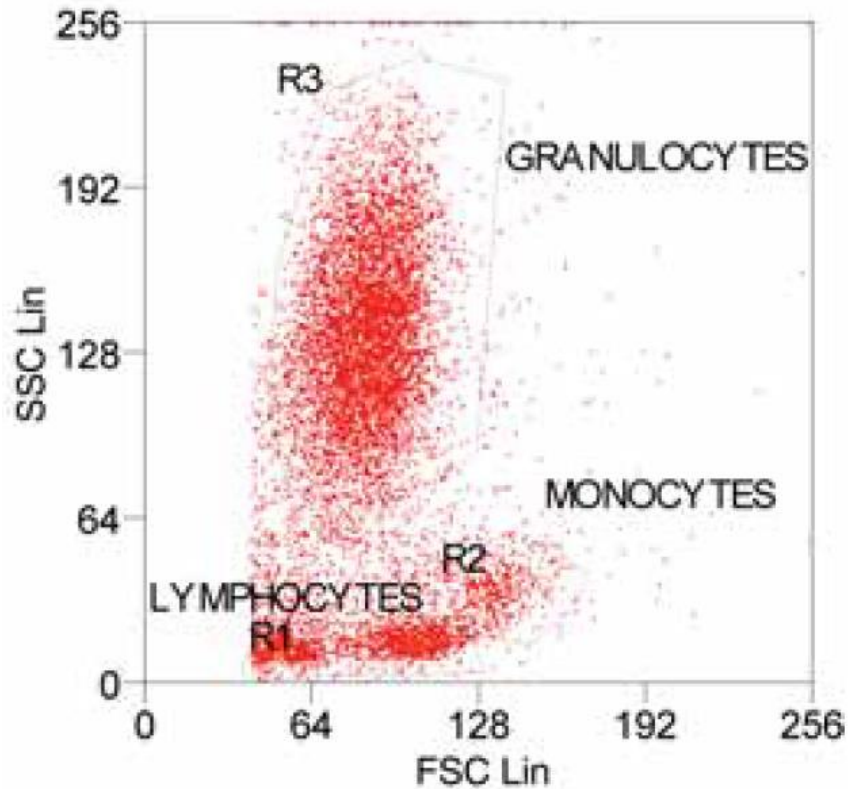
Density plot



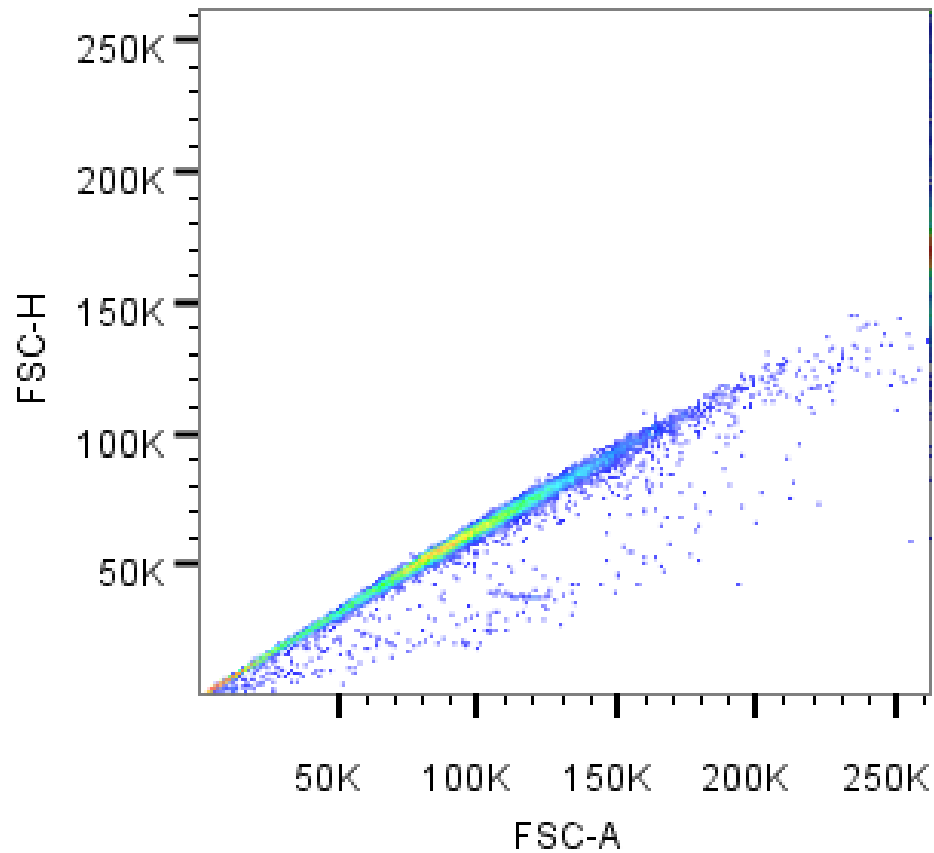
Contour diagram



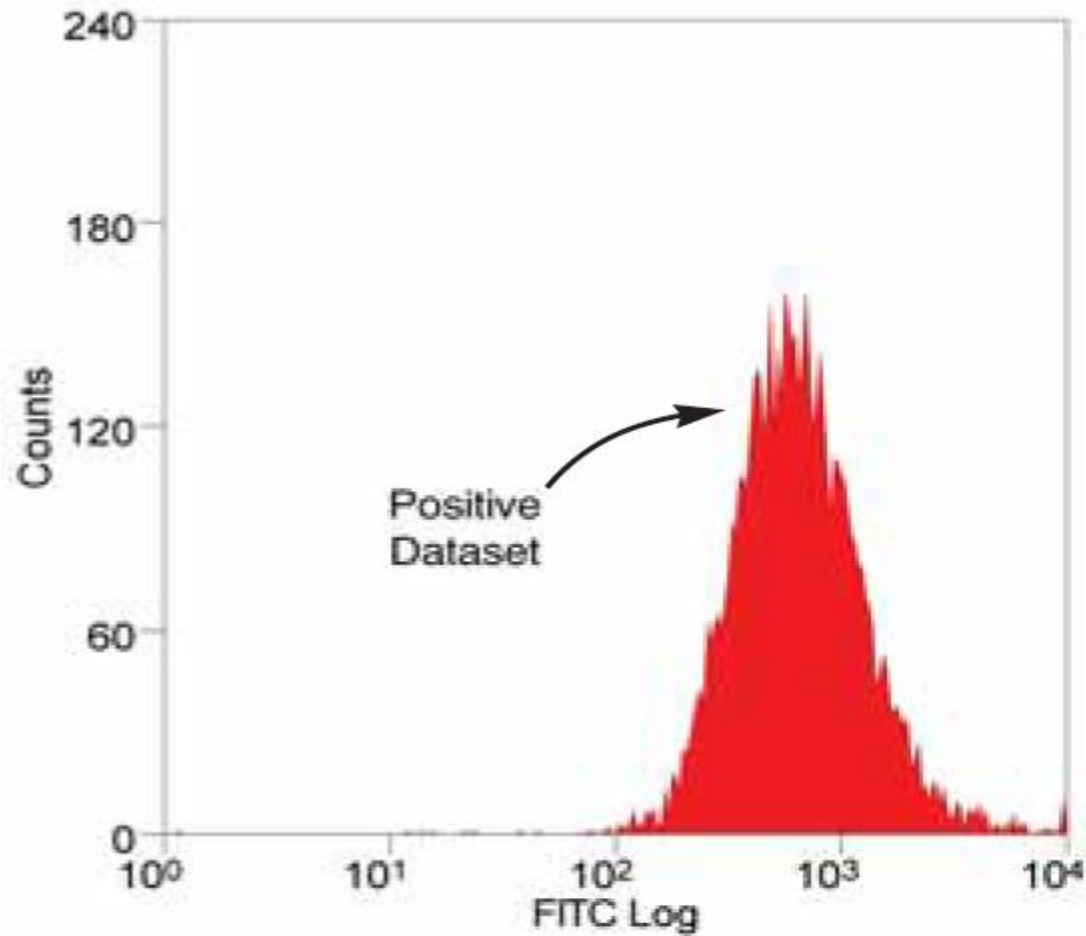
# Analysis of CD markers



# Discriminating doublets

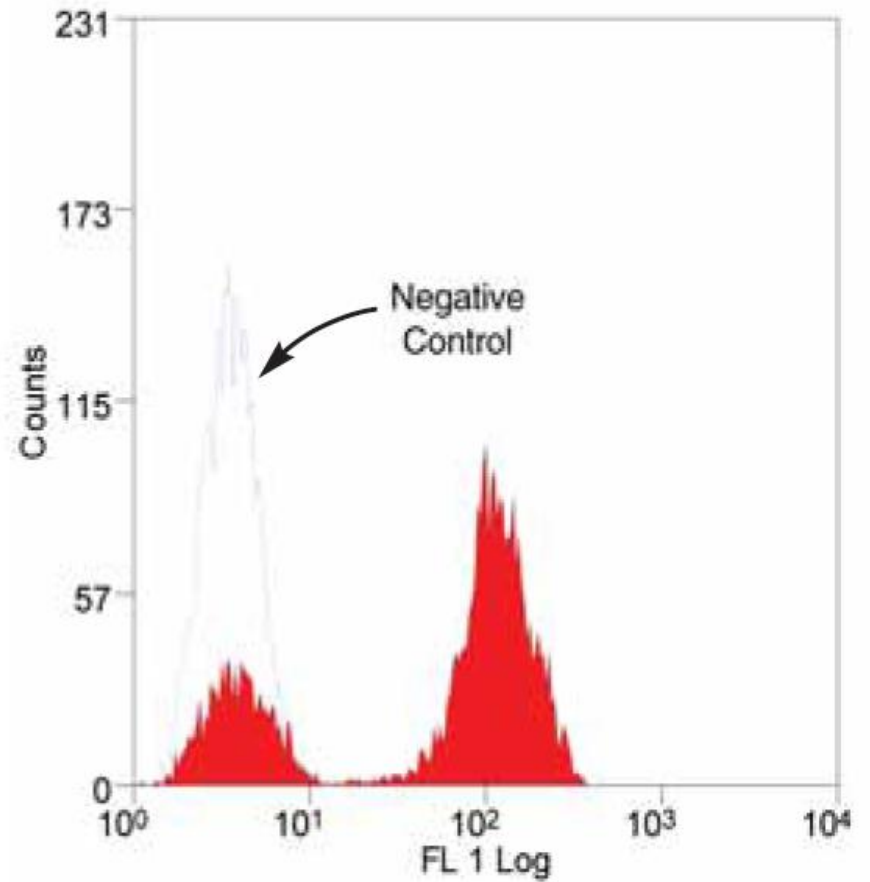
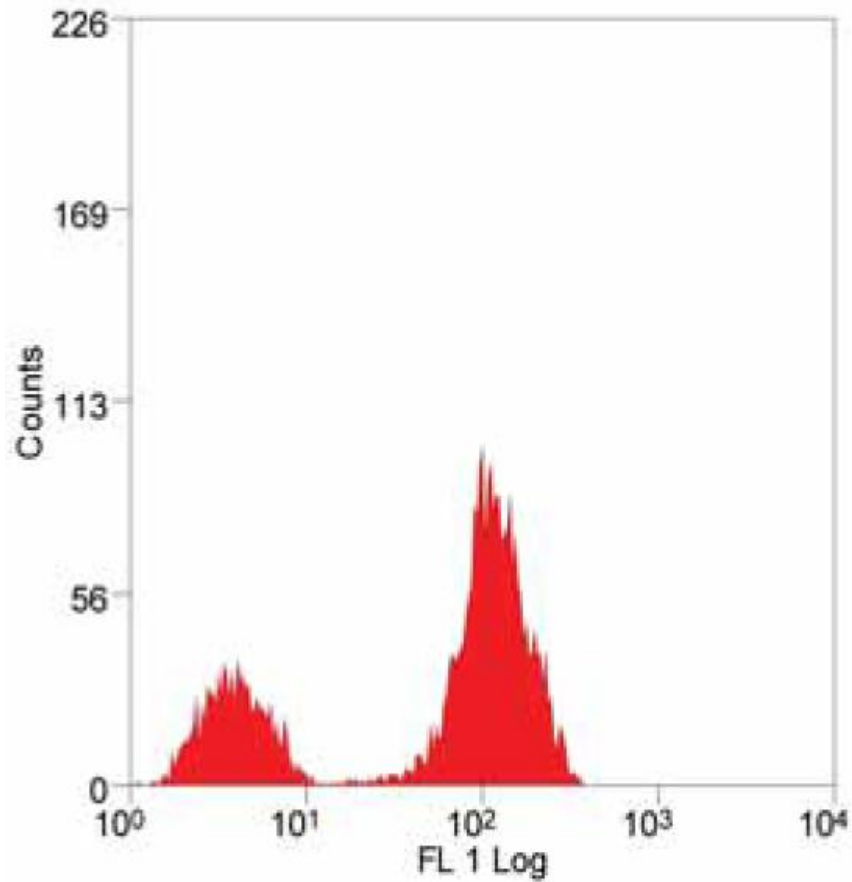


# Single parameter histogram

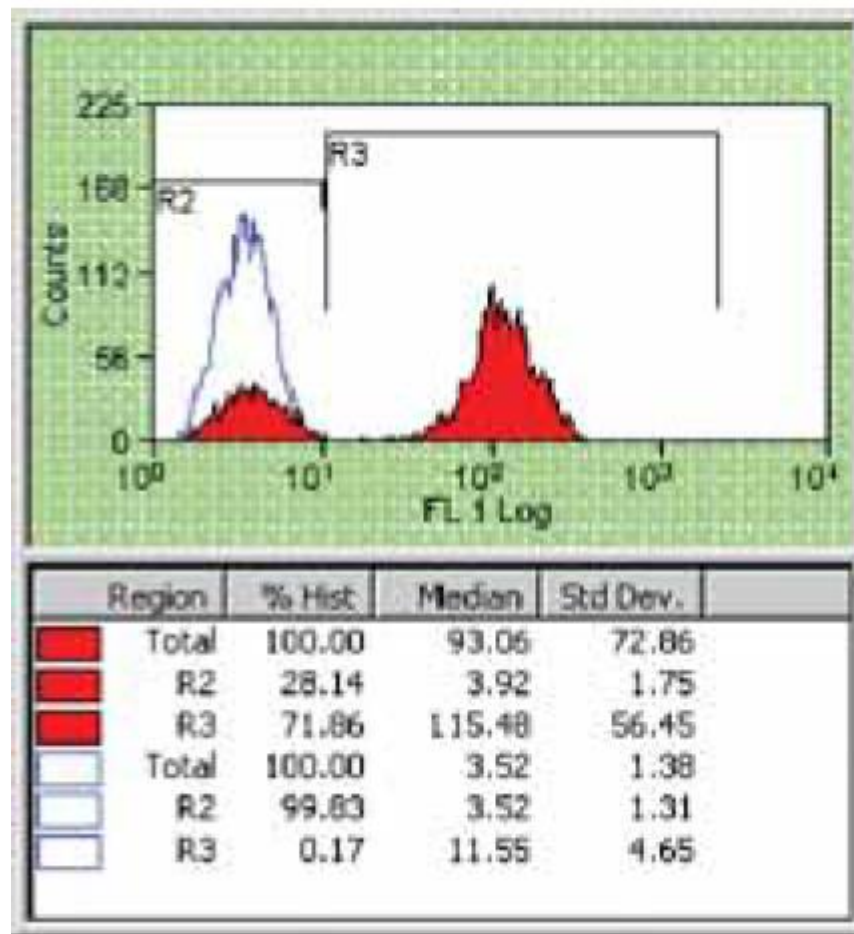




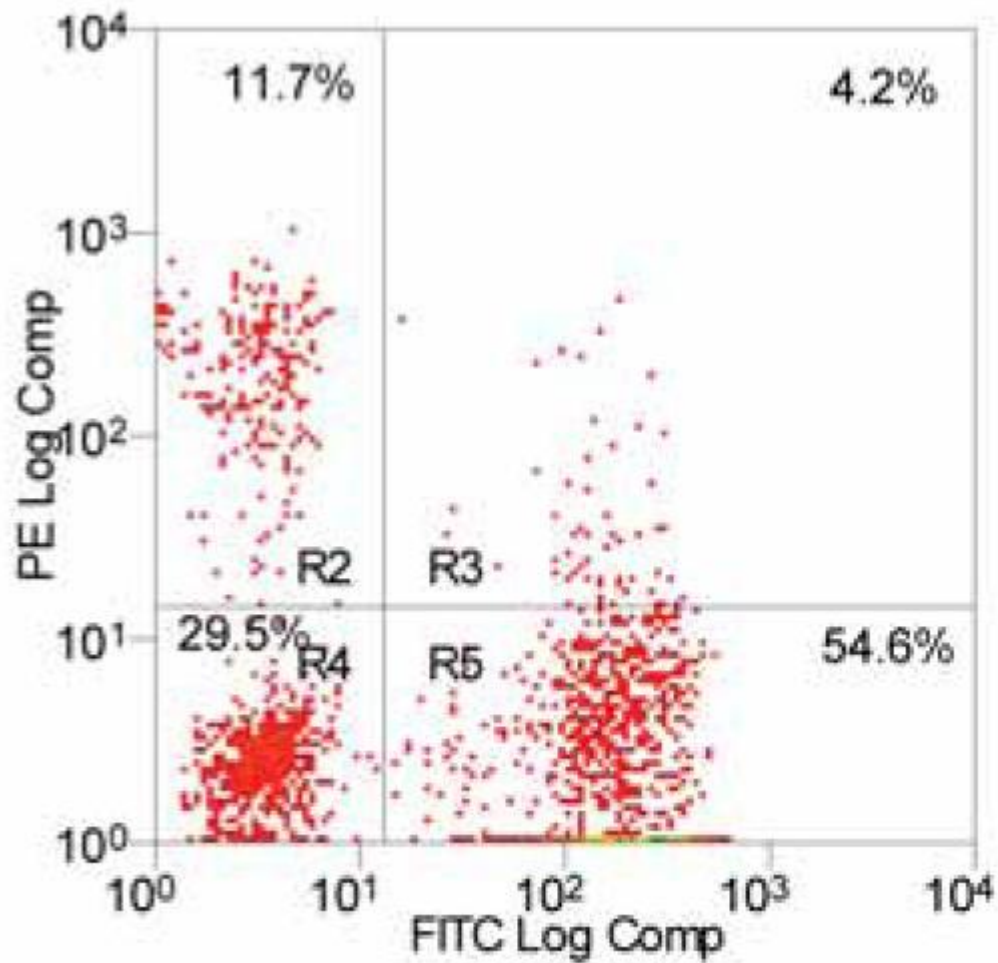
# Negative isotype control



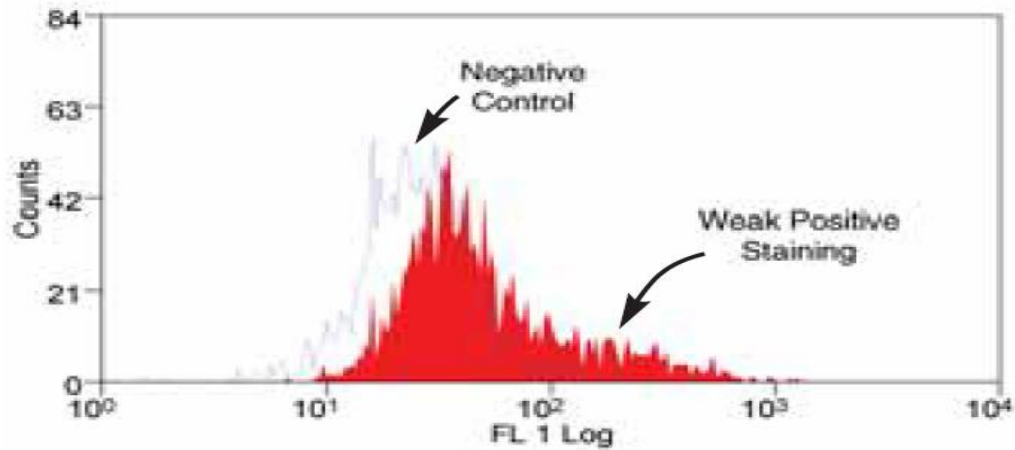
# Statistical analysis



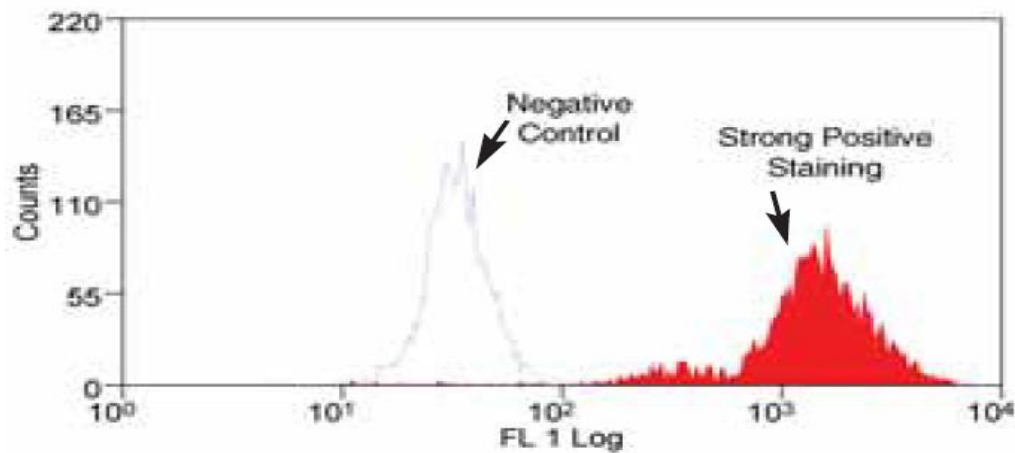
# Dual color histogram



# Intracellular markers



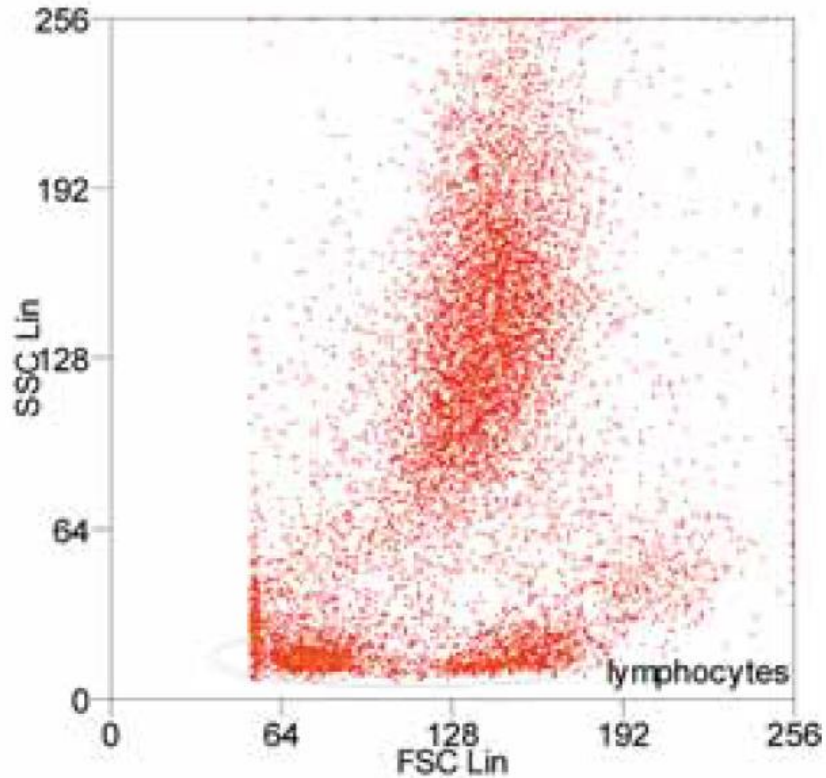
(a) BEFORE



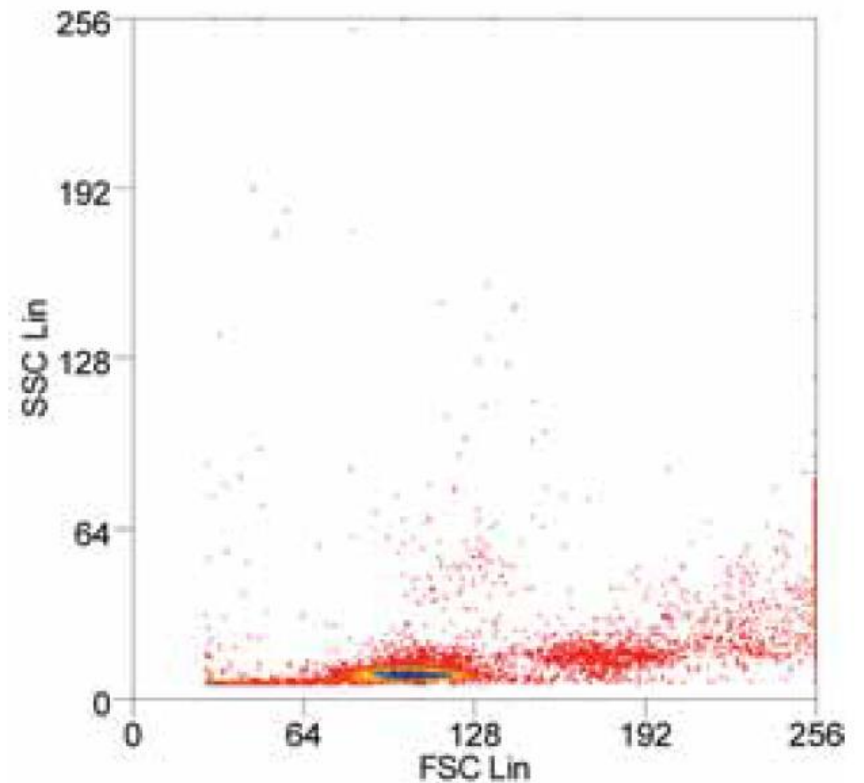
(b) AFTER

# Cell population shift

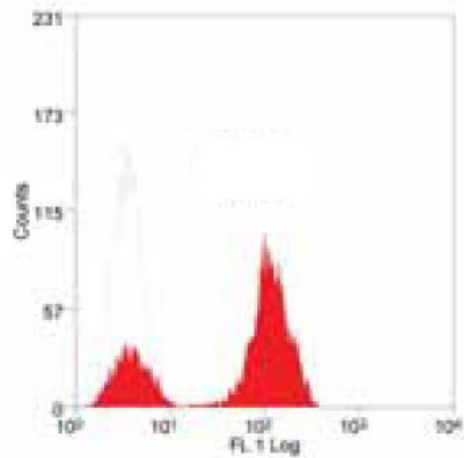
Normal person



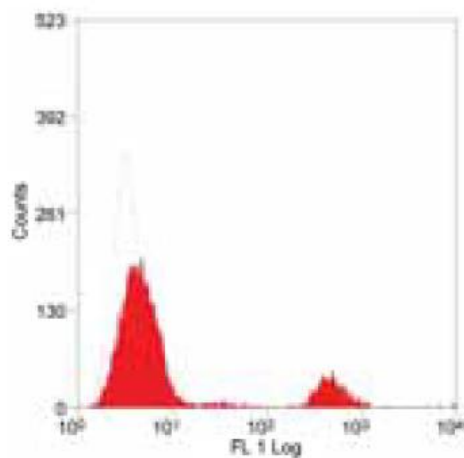
Leukemia patient



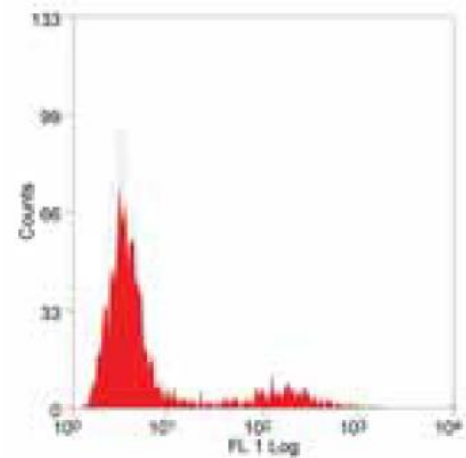
## Normal person



CD3

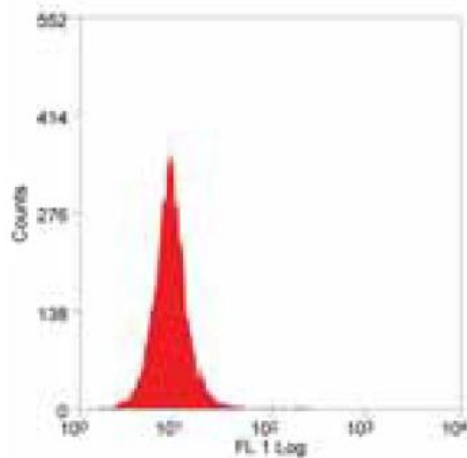


CD20

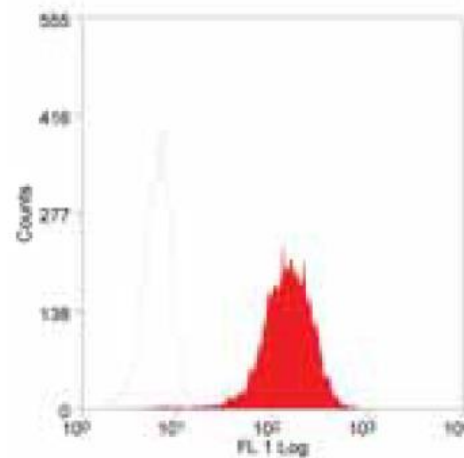


HLA-DR

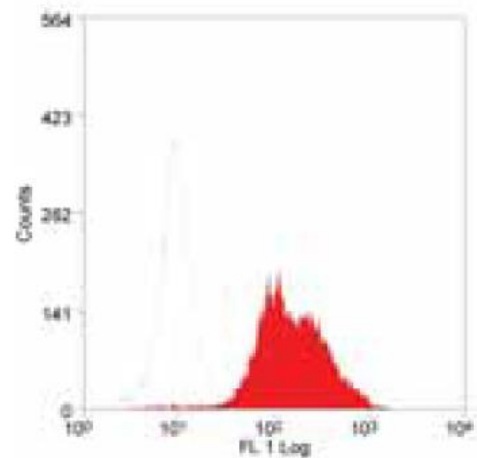
## Leukemia patient



CD3

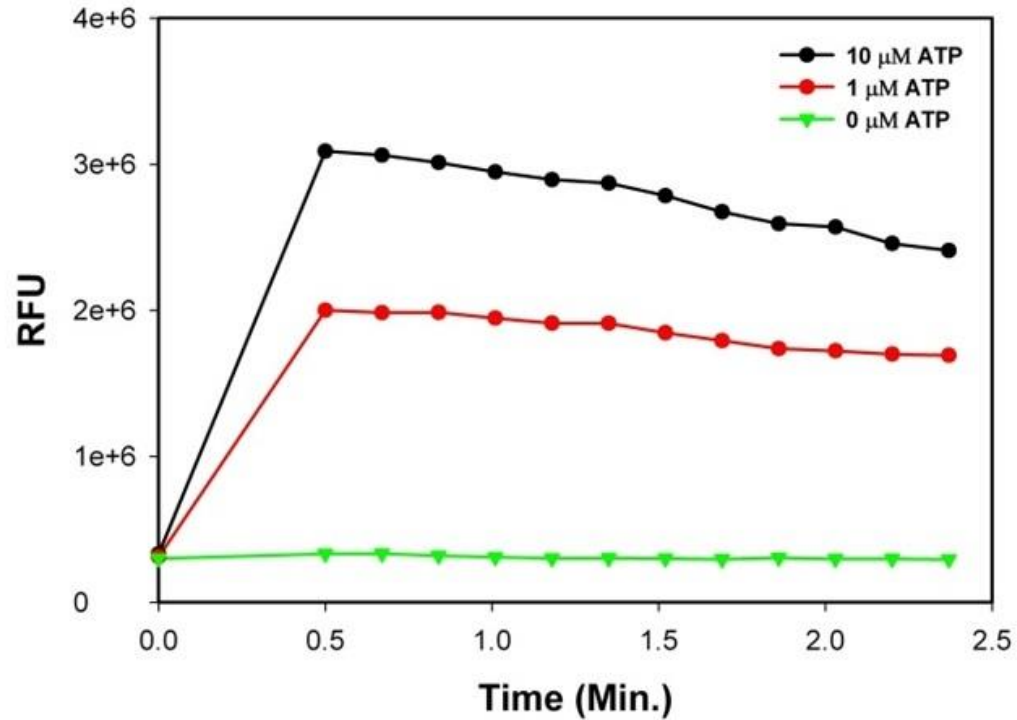
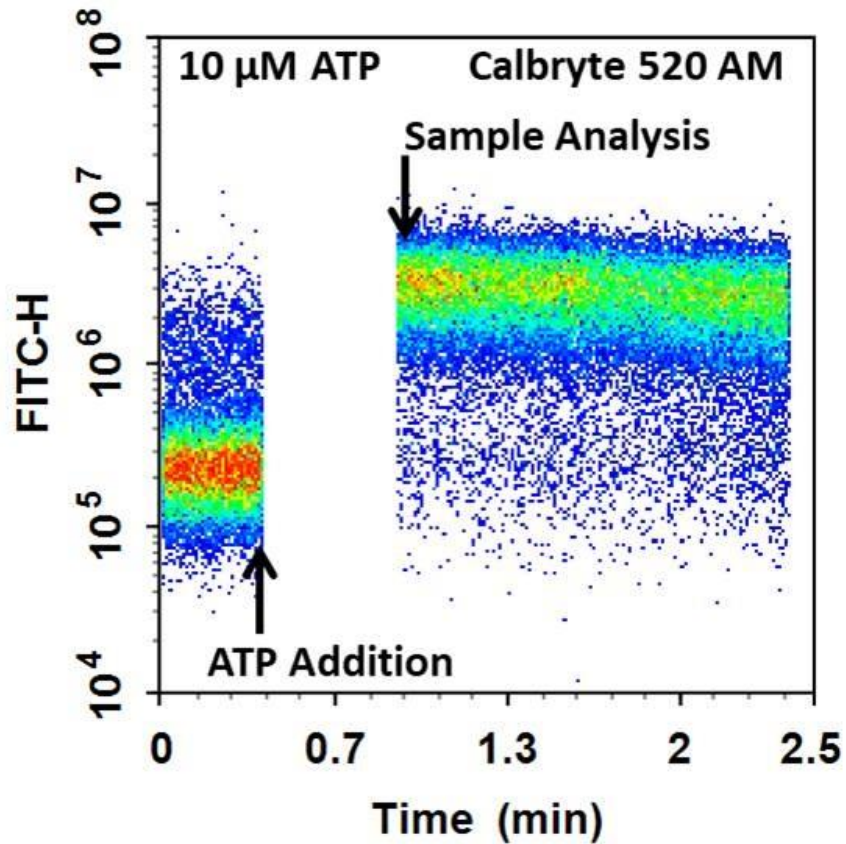


CD20

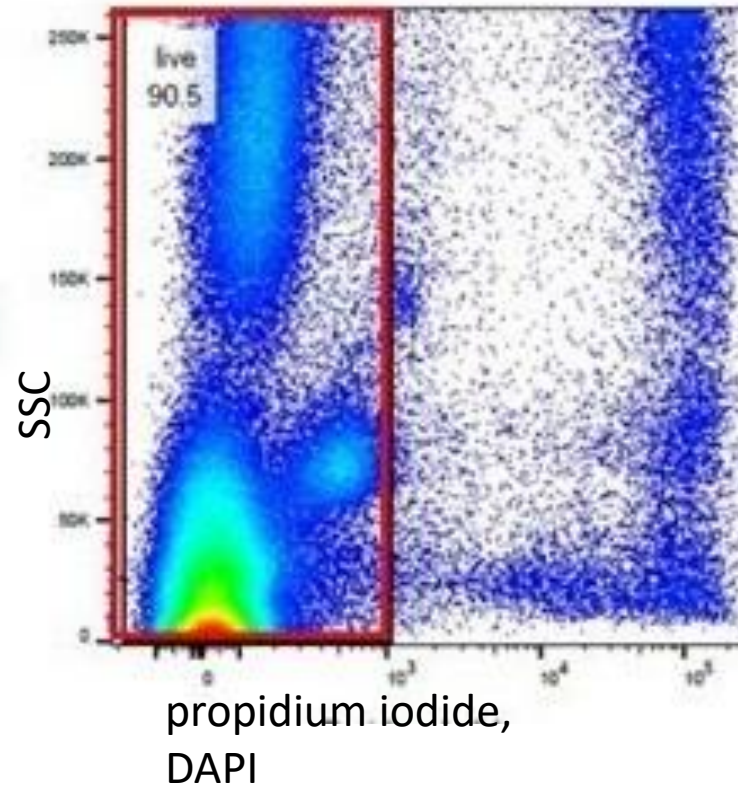


HLA-DR

# Calcium flux assay

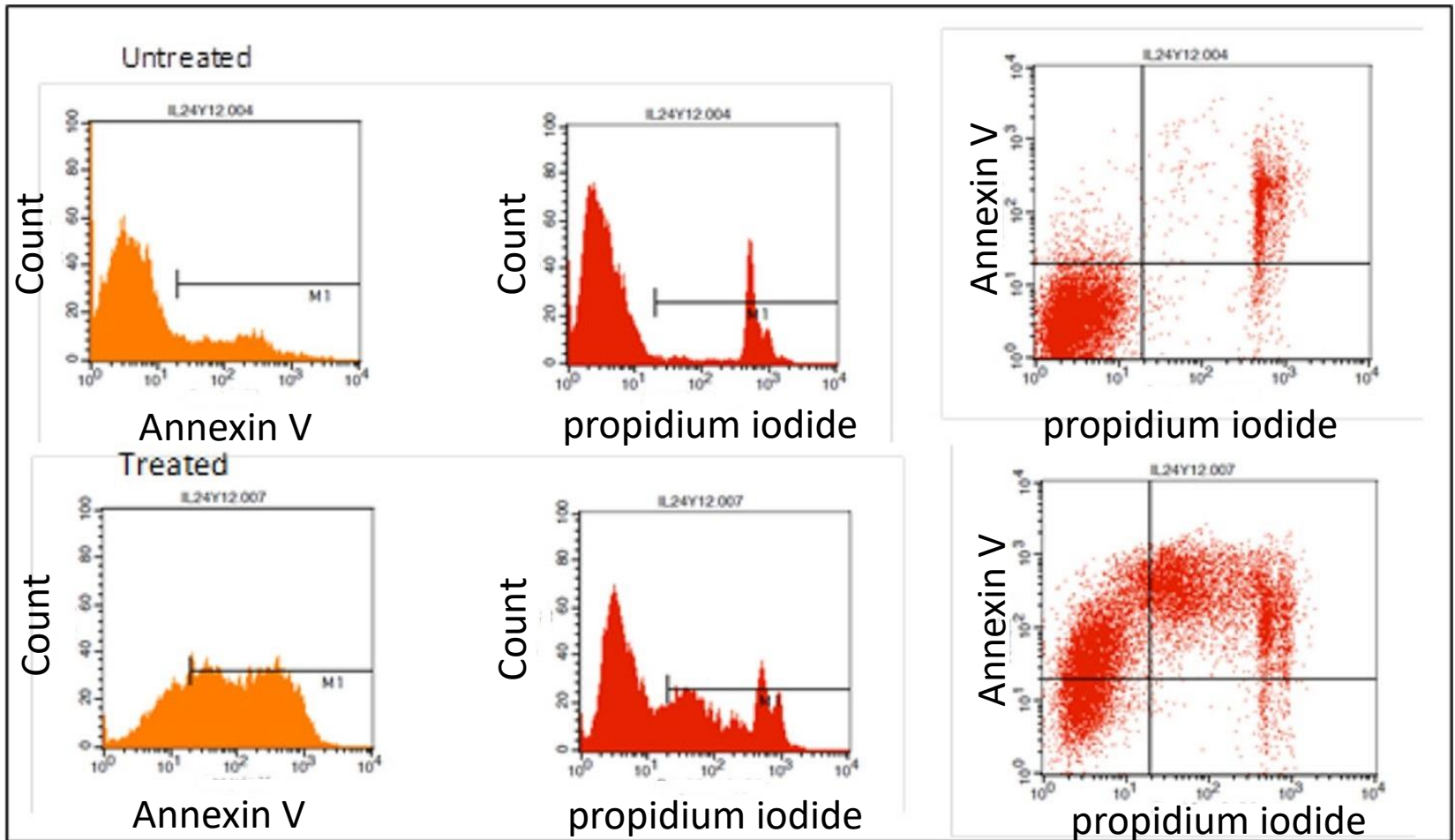


# Live and dead cells assay





# Apoptotic and necrotic cells



Thank You